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Wordom: a program for efficient analysis of molecular dynamics simulations

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ABSTRACT

Summary: Wordom is a versatile program for manipulation of molecular dynamics trajectories and efficient analysis of simulations. Original tools in Wordom include a procedure to evaluate significance of sampling for principal component analysis as well as modules for clustering multiple conformations and evaluation of order parameters for folding and aggregation. The program was developed with special emphasis on user-friendliness, effortless addition of new modules and efficient handling of large sets of trajectories.

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1 INTRODUCTION

Molecular dynamics (MD) simulations can produce several terabytes of data (Rueda *et al.*, 2007) whose manipulation and analysis is problematic in the absence of proper tools. As an example, sampling in the millisecond time-scale generates on the order of 10^7 to 10^8 coordinate frames (Settanni *et al.*, 2005). In this communication, a program is presented that can effectively access MD trajectory data and make them available for conversion, manipulation and analysis by different modules. In this way, it is possible to implement new procedures by focusing on the relevant equations and algorithms without having to deal with the data access module. The program provides general tools for analysis as well as special purpose modules which are not available in other programs (e.g. the efficient clustering of large sets of conformations and the evaluation of order parameters for monitoring polypeptide aggregation).

2 DATA MANIPULATION AND OUTPUT

Wordom can perform simple conversion and manipulation tasks, such as the extraction of one or more snapshots from a CHARMM (Brooks *et al.*, 1983), GROMACS (van der Spoel

et al., 2005), or NAMD (Kalé *et al.*, 1999) trajectory into a pdb (Berman *et al.*, 2000) or crd file (Brooks *et al.*, 1983), adding a single coordinate set from a pdb or crd file to a trajectory, merging segments of trajectories, calculating the average structure along a trajectory, extracting xyz coordinates of selected atoms, showing/modifying the content of a trajectory file header, etc. For most of these tools an atom selection mechanism is implemented. Notably, all of these functions are accessible with a single command line. Moreover, the coordinate and trajectory files generated by Wordom can be visualized by widely distributed graphics packages, e.g. VMD (Humphrey *et al.*, 1996) and Pymol (DeLano, 2002).

3 ANALYSIS OF MD SIMULATIONS

Wordom provides modules for the analysis of geometrical features (distances, contacts, dihedrals, hydrogen bonds), as well as evaluation of radius of gyration, DRMS (distance root mean square) and RMSD (root mean square deviation) from a reference structure. RMSD can also be computed with respect to the preceding structure, to highlight conformational changes along the trajectory. More complex types of analysis include a principal component analysis (PCA) module, the computation of quasi-harmonic entropy, several clustering algorithms and the evaluation of order parameters to monitor folding (i.e. number of native and non-native contacts) and amyloid aggregation.

3.1 PCA analysis

PCA is a useful technique for separating large amplitude motions from irrelevant fluctuations along an MD trajectory (Amadei *et al.*, 1993). The approach consists of diagonalizing the covariance matrix of atomic fluctuations with respect to the average structure, thus obtaining a set of eigenvectors and corresponding eigenvalues which represent the axes of maximal variance in the protein motion. An original feature in the PCA module is a procedure for iterative PCA evaluation along trajectory intervals of increasing length (e.g. snapshots 1–1000, snapshots 1–2000, etc.) to evaluate the significance of sampling. The eigenvectors computed at each time interval are projected into the ones of the preceding (i.e. shorter) time interval. A value close to 1 along the diagonal of the projection matrix indicates convergence of the corresponding eigenvector(s) suggesting that trajectory elongation is not necessary for the

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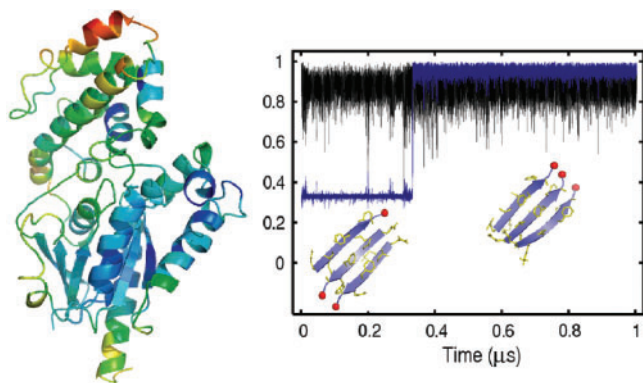


Fig. 1. (Left) Ribbon diagram of the α -subunit of protein Gq colored according to the length (red: large; green: intermediate; blue, small) of the atomic component of the first eigenvector on a 6-ns MD run by the Wordom PCA tool and displayed by Pymol (DeLano, 2002). (Right) Time series of P_1 (blue) and P_2 (black) (Cecchini *et al.*, 2004) along an MD trajectory of three Q15KLVFFA₂₁ heptapeptides from the Alzheimer's A β -peptide. Insets show representative snapshots of mixed and parallel β -sheet aggregates corresponding to low and high values of P_1 , respectively.

calculation of those eigenvectors. The PCA module in Wordom can also be used to generate pdb files where the last field of each atom entry (which usually contains the crystallographic β -factor) is set equal to the length of the vector defined by the components of the selected eigenvector along the corresponding atom. This quantitative information on the protein segment(s) involved in the most relevant concerted motion can be displayed directly with available graphics programs (Fig.1, left).

3.2 Quasi-harmonic entropy

The calculation of the quasi-harmonic entropy is based on the diagonalization of the mass-weighted covariance matrix of the atomic fluctuations (Andricioaei and Karplus, 2001). The $3n - 6$ non-zero eigenvalues λ_i are related to the quasi-harmonic frequencies $\omega_i = \sqrt{kT/\lambda_i}$ (where T is the simulation temperature and k the Boltzmann constant), which in turn yield the vibrational entropy $S = k \sum_i^{3n-6} \left(\frac{\hbar\omega_i/kT}{e^{\hbar\omega_i/kT} - 1} - \ln(1 - e^{-\hbar\omega_i/kT}) \right)$, the quasi-harmonic vibrational energy $E = \sum_i^{3n-6} \left(\frac{\hbar\omega_i}{2} + \frac{\hbar\omega_i}{e^{\hbar\omega_i/kT} - 1} \right)$ and the vibrational specific heat $C_V = \partial E / \partial T$. The entropy measured in this way is directly related to the conformational space explored by the system along the MD simulation and provides a wide range of applications from estimation of the entropic effects of mutations in the native state of proteins to the determination of the relative entropy of ligands in a binding pocket (Thorpe and Brooks, 2004).

3.3 Clustering algorithms

Clustering can be performed using two metrics (DRMS or RMSD) and three algorithms: hierarchical (Johnson, 1967), quality threshold (Heyer *et al.*, 1999) or leader-like (Hartigan, 1975). These algorithms have different CPU-time requirements and yield different clustering performances, hierarchical and quality threshold being slower but more precise and leader-like being much faster but dependent on the frame order. For the

former, it is possible to first take into account a subset of the snapshots and then cluster the remaining snapshots to reduce memory and CPU-time requirements. In the case of RMSD-based clustering, superposition can be carried out before computing the distance, depending on whether the relative position of the structures is relevant. It is also possible to superpose the structures according to a subset of atoms and cluster them relatively to another subset, which is useful for analyzing multiple poses obtained by ligand docking into a single protein structure.

3.4 Order parameters

Wordom efficiently evaluates the number of contacts, i.e. α -pairs with a distance below a given threshold (input value). The subset of contacts which are present in the native structure is a useful variable to monitor the progress of (un)folding (Chan and Dill, 1998).

The polar and nematic order parameters are used to study amyloid formation (Cecchini *et al.*, 2004). They are defined as $P_1 = \frac{1}{N} \sum_{i=1}^N \hat{\mathbf{z}}_i \cdot \hat{\mathbf{d}}$ and $P_2 = \frac{1}{N} \sum_{i=1}^N \frac{3}{2} (\hat{\mathbf{z}}_i \cdot \hat{\mathbf{d}})^2 - \frac{1}{2}$, where $\hat{\mathbf{d}}$ is a unit vector pointing in the preferred direction of alignment, $\hat{\mathbf{z}}_i$ is a suitably defined molecular vector and N is the number of peptides. These order parameters capture different orientational properties of the system and yield useful and complementary information. The polar P_1 describes the polarity of the system, i.e. how much the molecular vectors ($\hat{\mathbf{z}}_i$) point in the same direction, and discriminates between parallel, antiparallel or mixed ordered aggregates. The nematic P_2 describes the orientational order of the system and discriminates between ordered and disordered conformations. The evaluation of P_1 and P_2 facilitates the analysis of simulations of peptide aggregation (Cecchini *et al.*, 2004) by measuring the degree of structural order during the fibril formation process (Fig.1, right).

4 CONCLUSIONS

Analyzing the ever growing amounts of MD trajectory data can become the rate-limiting step in MD simulation studies. The Wordom program facilitates this task. Manipulations of multiple trajectory files and simple geometric analyses require a single line of input, while input files for clustering and PCA analysis consist of about five lines. Therefore, for efficiently extracting information from massive sets of MD data Wordom is better suited than other programs which require complicated input files and/or the use of a graphical interface. Moreover, thanks to low-level data access functions, adding new analysis modules requires only the coding of the core function allowing for a steadily growing library of algorithms introduced by the community of users.

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